

Nonpeptide $\alpha_v\beta_3$ antagonists. Part 10: In vitro and in vivo evaluation of a potent 7-methyl substituted tetrahydro-[1,8]naphthyridine derivative

Michael J. Breslin,^{a,*} Mark E. Duggan,^a Wasyl Halczenko,^a George D. Hartman,^a Le T. Duong,^b Carmen Fernandez-Metzler,^c Michael A. Gentile,^b Donald B. Kimmel,^b Chih-Tai Leu,^b Kara Merkle,^c Thomayant Prueksaritanont,^c Gideon A. Rodan,^b Sevgi B. Rodan^b and John H. Hutchinson^{a,†}

^aDepartment of Medicinal Chemistry, Merck Research Laboratories, West Point, PA 19486, USA

^bDepartment of Bone Biology and Osteoporosis Research, Merck Research Laboratories, West Point, PA 19486, USA

^cDepartment of Drug Metabolism, Merck Research Laboratories, West Point, PA 19486, USA

Received 11 February 2004; accepted 14 June 2004

Available online 17 July 2004

Abstract—Subtle modifications were incorporated into the structure of clinical candidate **1**. These changes were designed to maintain potency and selectivity while inducing changes in physical properties leading to improved pharmacokinetics in three species. This approach led to the identification of **4** as a potent, selective $\alpha_v\beta_3$ receptor antagonist that was selected for clinical development based on an improved PK profile and efficacy demonstrated in an in vivo model of bone turnover.

© 2004 Elsevier Ltd. All rights reserved.

Osteoporosis is a disease most commonly found in post-menopausal women, where the combination of estrogen deficiency and aging results in the loss of a significant portion of the peak bone mass attained as a young adult. The characteristic skeletal fragility and high incidence of fracture are due to a decrease in mechanical bone strength resulting from an imbalance between the activities of bone-resorbing osteoclast cells and bone-depositing osteoblast cells.^{1–4} One potential approach to combatting this disease is to reduce the rate of osteoclast-mediated bone resorption by antagonism of the integrin $\alpha_v\beta_3$.⁵ $\alpha_v\beta_3$ is highly expressed in osteoclasts and is thought to be involved in the adhesion, activation, and migration of osteoclasts on the bone surface as well as osteoclast polarization.^{6,7} Several proteins that possess the three-amino acid sequence arginine–glycine–aspartic acid (RGD), including osteopontin, bone sialoprotein, vitronectin, and fibrinogen, bind with high affinity to $\alpha_v\beta_3$.⁸ Antibodies to $\alpha_v\beta_3$, RGD-peptides, and

small molecule mimetics of RGD have been shown to be efficacious in in vitro and in vivo models of bone resorption.⁹ These results suggest that antagonism of the RGD binding domain of $\alpha_v\beta_3$ by high affinity nonpeptidic RGD mimetics may result in a treatment for osteoporosis.

Previous reports from this laboratory have described the chain-shortened imidazolidinone **1** as an orally bioavailable zwitterionic antagonist that was selected for evaluation in human clinical trials.¹⁰ Compound **1** was projected to display a pharmacokinetic profile in humans to support twice daily oral dosing. Satisfied with the in vitro potency and selectivity of **1**, we sought to identify a related compound with improved pharmacokinetics that could be effective in humans following once-a-day administration. Herein, we describe modifications to the *N*-terminus of **1** that were designed to increase lipophilicity and block a potential site of human metabolism. These efforts resulted in compound **4**, which showed an enhanced pharmacokinetic profile and was chosen for clinical development.

Compound **1** was selected as a clinical candidate for the prevention and treatment of osteoporosis based on

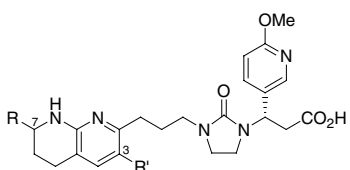
* Corresponding author. Tel.: +1-215-652-7928; fax: +1-215-652-7310; e-mail: michael_breslin@merck.com

† Present address: Merck & Co., Inc. MRLSDB2, 3535 General Atomics Court, San Diego, CA 92121, USA.

excellent in vitro potency and selectivity, a good pharmacokinetic profile and efficacy shown in three different in vivo models of bone resorption.¹⁰ After extensive SAR studies around aryl substituents on the β -alanine C-terminus of the imidazolidinone core series, 2-methoxy-pyridine was found to possess a desirable balance between basicity, polarity, and the degree of binding to human plasma proteins.^{11,12} Typically, polar compounds in the imidazolidinone series, as defined by a log *P* value approaching zero, suffered from poor pharmacokinetic profiles due to low bioavailability. Lipophilic compounds with a log *P* value approaching 1.0 usually displayed good oral bioavailability and half-life, but high protein binding reduced in vivo efficacy due to low free drug concentration.

We sought to introduce conservative modifications to the *N*-terminus of **1** that would not adversely alter the in vitro potency and selectivity, but would induce favorable changes in physical properties. The methyl group was chosen as a small substituent that could enhance lipophilicity without having a dramatic effect on free drug concentration. In a related series, 7-methyl substitution on the tetrahydro-[1,8]naphthyridine (THN) ring was found to be 2-fold more potent than 6-methyl substitution. In this series, substitution at the C-3 position of the THN ring was extensively studied showing that electron withdrawing groups decreased binding affinity while electron donating groups increased potency.¹³ Substitution on the unsaturated ring of the THN may largely affect the binding affinity and physical properties of the *N*-terminus by modulation of the *pK_a*. When 3-methyl substitution was employed in the imidazolidinone series, the substitution had the desired effect on in vitro potency and physical properties. Compound **2** possessed comparable potency to **1** while having less protein binding in spite of being more lipophilic (Table 1). This substitution had detrimental effects on the pharmacokinetic profile of **2**; clearance increased dramatically while half-life decreased. Substitution with slightly larger aliphatic groups such as ethyl caused a 10–20-fold loss of selectivity versus the related integrin $\alpha_v\beta_5$ when compared with **1** and **2**.

Table 1. In vitro binding affinity and physical properties of THN substituted $\alpha_v\beta_3$ antagonists



Compd	R	R'	SPAV3 ^a IC ₅₀ (nM)	PB (%) ^b	Log <i>P</i>
1	H	H	0.08	88	0.21
2	H	CH ₃	0.13	78	0.62
3	(<i>S</i>)-CH ₃	H	0.11	92	0.56
4	(<i>R</i>)-CH ₃	H	0.11	95	0.69

^a Binding to the $\alpha_v\beta_3$ receptor using a scintillation-proximity assay (SPAV3), *n* = 235 for **1**, *n* = 2 for **2** and **3**, *n* = 7 for **4**.

^b Human plasma protein binding.

Although in vitro metabolism studies on **1** using liver microsomes from several species showed minimal metabolism, the C-7 methylene position of the THN ring was identified as a site of oxidative metabolism. The C-7 hydroxylated metabolite could undergo further oxidation to yield a lactam or eliminate and aromatize to give the fully unsaturated naphthyridine.¹⁰ We focused on placing a methyl substituent at the C-7 position of the THN ring knowing that this substitution was tolerated in a related series and that blocking a site of metabolism could improve pharmacokinetics. The first synthesis of a 7-methyl substituted THN ring in the imidazolidinone series yielded a racemic compound, which was separated by chiral chromatography to give **3** and **4**.¹⁴ Both **3** and **4** were equipotent in the $\alpha_v\beta_3$ binding assay (SPAV3) and their physical properties were similar.¹⁵ Later asymmetric synthesis established the absolute stereochemistry at the C-7 methyl position to be *S* for compound **3** and *R* for compound **4**. We concentrated further studies on diastereomer **4**.

Like compound **1**, **4** displayed good selectivity against several related integrins. Compound **4** displayed a 50-fold selectivity for $\alpha_v\beta_3$ over $\alpha_v\beta_5$ as demonstrated by an IC₅₀ of 5 nM (*n* = 7) in an $\alpha_v\beta_5$ binding assay.¹⁴ Affinity for the fibrinogen receptor $\alpha_{IIb}\beta_3$, as measured by a platelet aggregation assay, was found to be weak with an IC₅₀ > 10 μ M.¹⁶ While the addition of the C-7 methyl group on the THN ring did not adversely alter the vitro potency and selectivity, the physical properties of **4** were changed. As compared to compound **1**, Log*P* increased favorably from 0.21 to 0.69 while protein binding increased slightly from 88% to 95%. This compound displayed improved pharmacokinetics in three species (Table 2). Clearance remained high in rats, but slightly improved over compound **1**. Half-life decreased slightly, but oral bioavailability doubled (50% vs 26%). The pharmacokinetic profile in dog and monkey was characterized by low clearance (1.7 and 3.5, respectively) and high oral bioavailability (83% and 75%) while *t*_{1/2} was unchanged from that observed for **1**. The pharmacokinetic profile of compound **4** was highlighted by reduced clearance, comparable half-life, and improved bioavailability.

On the basis of its excellent in vitro potency, selectivity, and improved pharmacokinetic profile, **4** was chosen for study in a rodent model of bone resorption. Compound **4** was administered in two different experiments to

Table 2. Pharmacokinetics in several species^a

Species	Compd	CL (mg/min/kg)	<i>T</i> _{1/2} (h)	<i>F</i> (%)
Rat	1	47	3	26
	4	41	1–2	50
Rhesus monkey	1	9.0	2	74
	4	3.5	2	75
Dog	1	6.4	3.5	64
	2	18	2.2	55
	4	1.7	4	83

^a Compounds dosed at 0.2 mpk iv and 1 mpk po in water.

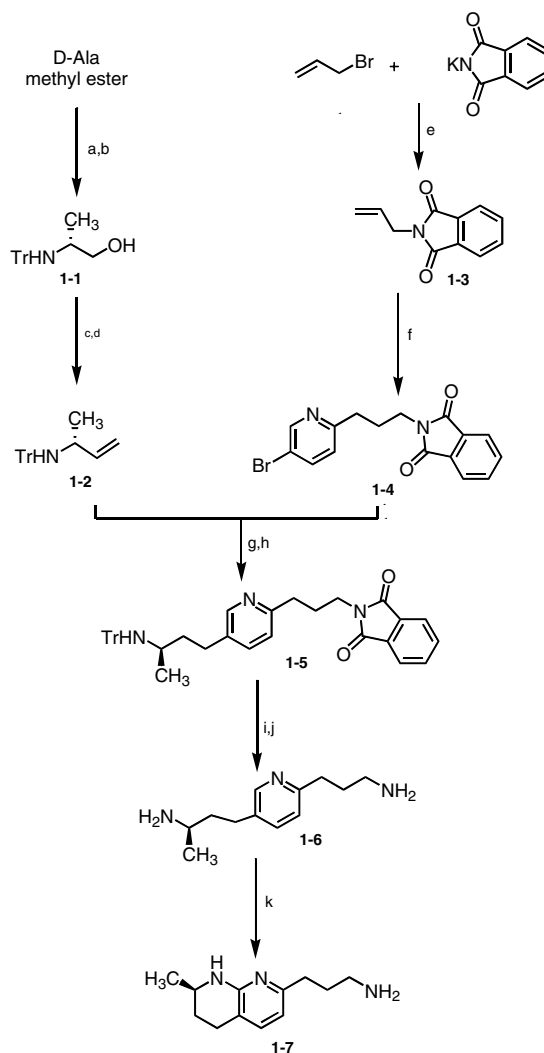
Table 3. Effect of **1** and **4** on growing young male rats

Compd	Free C_{ss} ^a (nM)	DFMBMD ^b (mg/cm ²)	% Increase over vehicle
Vehicle	—	112.5 ± 5.8	—
1	170	133.0 ± 7.5 ^c	+18
4	190	137.0 ± 10.0 ^c	+22
4	560	147.9 ± 7.3 ^{c,d}	+32
4	1440	161.0 ± 13.7 ^{c,d}	+43
Vehicle	—	118.6 ± 9.2	—
1	390	145.0 ± 10.0 ^c	+22
1	1550	157.2 ± 12.3 ^{c,e}	+33
4	170	141.1 ± 9.7 ^c	+19
4	1100	156.6 ± 11.8 ^{c,f}	+32

^a Free C_{ss} (nM)—free steady state serum concentration.^b DFMBMD—distal femoral metaphysis bone mineral density.^c >Vehicle ($P < 0.0001$).^d >190 nM **4** ($P < 0.0001$).^e >390 nM **1** ($P < 0.002$).^f >170 nM **4** ($P < 0.0001$).

young, rapidly growing male rats by minipump infusion over ten days (Table 3). Compound **1** was used as a positive control and at the end of the experiment bone mineral density at the distal femoral metaphysis was measured and compared to that of vehicle-treated animals. Both compounds produced a significant increase in distal femoral metaphysis bone mineral density (DFMBMD), which was significantly different than the vehicle control group. In the first experiment, compound **4** increased DFMBMD in an exposure-dependent fashion, where free steady state serum concentration (C_{ss}) varied from 190 to 1440 nM with a maximum effect at +43% above the vehicle. The DFMBMD increase achieved with **4** at 190 nM (+22%) was approximately the same as seen with **1** at 170 nM (+18%). In the second experiment, both **1** and **4** produced statistically significant, exposure-dependent increases in DFMBMD. The C_{ss} of **1** varied from 390 to 1550 nM with a maximum effect at +33% while a lower C_{ss} of **4** (170–1100 nM) was able to produce similar effects (maximum at +32%). These data suggest that **4** is at least as efficacious as **1** for inhibiting bone resorption in rats.

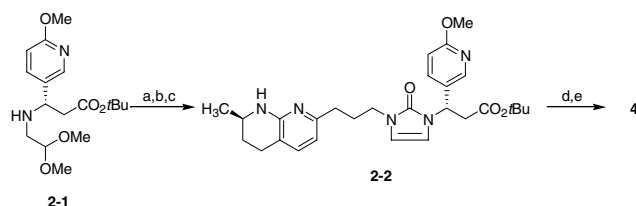
The asymmetric synthesis of **4** was accomplished using a three part convergent synthesis, which utilized the C-terminal intermediates developed for the process scale preparation of compound **1**.¹⁷ The synthesis of the N-terminus of **4** (Scheme 1) is accomplished in 10 steps from commercially available materials. D-Alanine methyl ester was trityl protected and reduced to the alcohol **1-1** with LAH. Oxidation by pyridine- SO_3 yielded the aldehyde, which was immediately used in a Wittig reaction with methyl triphenylphosphonium bromide to give **1-2** in 93% yield for four steps.¹⁸ Allyl bromide was coupled to potassium phthalimide by heating in DMF followed by cross-coupling to 2,5-dibromopyridine to give **1-4** in 50% yield. Heck coupling of **1-2** and **1-4** gave **1-5**, which was fully deprotected and subsequently cyclized via an intramolecular Chichibabin reaction to yield the key intermediate **1-7** in 60% yield for the last four steps.



Scheme 1. Reagents and conditions: (a) trityl chloride, Et_3N , CH_2Cl_2 ; (b) $LiAlH_4$, THF; (c) pyridine- SO_3 , Et_3N , DMSO, CH_2Cl_2 ; (d) methyl triphenylphosphonium bromide, $n-BuLi$, THF, 0 °C; (e) DMF, 70 °C; (f) 9-BBN, 2,5-dibromopyridine, K_2CO_3 , DPPF, $Pd(OAc)_2$, DMF; (g) 9-BBN, THF; (h) **1-4**, K_2CO_3 , DPPF, $Pd(OAc)_2$, DMF; (i) HCl, EtOAc; (j) hydrazine, EtOH, MTBE; (k) NaH, xylenes.

The C-terminal intermediate **2-1** was converted to the chlorocarbamate with triphosgene and then reacted directly with **1-7** to give an unsymmetric urea that was cyclized to the imidazoline-2-one **2-2** by the addition of 2 M sulfuric acid to the reaction mixture.¹⁷ Catalytic hydrogenation smoothly reduced the olefin and this was followed by hydrolysis of the ester with 6 M sulfuric acid to give **4** (Scheme 2). The original chiral resolution of **3** and **4** was performed on the *tert*-butyl ester following hydrogenation. Analysis of the *tert*-butyl ester of **4** showed 98.8% de was obtained, which is comparable to the ee of the D-alanine methyl ester starting material.

In summary, we have incorporated subtle changes to the structure of clinical development candidate **1** to effect changes in physical properties and improve pharmacokinetics. A strategically placed methyl group on the N-terminus of the molecule was found to slightly increase lipophilicity without dramatically increasing



Scheme 2. Reagents and conditions: (a) triphosgene, Et₃N, THF; (b) 1–7, Et₃N, THF, 40 °C; (c) 2 M H₂SO₄; (d) Pd(OH)₂, EtOH, AcOH, H₂ (70 psi); (e) 6 M H₂SO₄.

protein binding. On the basis of an improved pharmacokinetic profile in three species and efficacy demonstrated in an in vivo model of bone turnover, **4** was selected for clinical development.

References and notes

- Eastell, R. N. *Engl. J. Med.* **1998**, *338*, 736.
- Graul, A. I.; Sorbera, L. A.; Prous, J. R. *Drugs Today* **2001**, *37*, 703.
- Ershler, W. B. *Front. Biomed.* **2000**, *41*.
- Sato, M.; Grese, T. A.; Dodge, J. A.; Bryant, H. U.; Turner, C. H. *J. Med. Chem.* **1999**, *42*, 1.
- Rodan, S. B.; Rodan, G. A. *J. Endocrinol.* **1997**, *154*, S47.
- Vaananen, H. K.; Liu, Y.-K.; Lehenkari, P.; Uemara, T. *Mater. Sci. Eng. C* **1998**, *6*, 205.
- Shankar, G.; Horton, M. *Adv. Organ. Biol.* **1998**, *5B*, 315.
- Duong, L. T.; Rodan, G. A. *Front. Biosci.* **1997**, *3*, d757.
- For reviews see: (a) Miller, W. H.; Keenan, R. M.; Willette, R. N.; Lark, M. W. *Drug. Discov. Today* **2000**, *5*, 397; (b) Hartman, G. D.; Duggan, M. E. *Exp. Opin. Invest. Drugs* **2000**, *9*, 1281; (c) Duggan, M. E.; Hutchinson, J. H. *Exp. Opin. Ther. Pat.* **2000**, *10*, 1367; (d) Coleman, P. J.; Duong, L. T. *Exp. Opin. Ther. Pat.* **2002**, *12*, 1009.
- Hutchinson, J. H.; Halczenko, W.; Brashear, K. M.; Coleman, P. J.; Duong, L. T.; Fernandez-Metzler, C.; Gentile, M. A.; Fisher, J. E.; Hartman, G. D.; Huff, J. R.; Kimmel, D. B.; Leu, C.; Meissner, R. S.; Merkle, K.; Nagy, R.; Pennypacker, B.; Perkins, J. J.; Prueksaritanont, T.; Rodan, G. A.; Varga, S. L.; Wesolowski, G. A.; Zartman, A. E.; Rodan, S. B.; Duggan, M. E. *J. Med. Chem.* **2003**, *46*, 4790.
- LogP: Measured from octanol, pH 7.4 buffer.
- Protein binding: Human plasma binding measured in a high-throughput manner using ultrafiltration of diluted plasma in a pH 7.4 buffer. The analysis is conducted at room temperature, and solutions are analyzed by HPLC.
- Wang, J.; Breslin, M. J.; Coleman, P. J.; Duggan, M. E.; Hunt, C. A.; Hutchinson, J. H.; Leu, C.; Rodan, S. B.; Rodan, G. A.; Duong, L. T.; Hartman, G. H. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 1049.
- Merck and Co. US Patent 6,472,403 B2, 2002.
- SPAV3 is a scintillation-proximity assay that measures the displacement of 4-[2-(2-aminopyridin-6-yl)ethyl]benzoyl-2(S)4-¹²⁵Iodophenylsulfonfylamino-β-alanine from purified human recombinant α_vβ₃. A similar SPA assay (SPAV5) utilizes the same ligand to measure its displacement from α_vβ₃. For protocol, see: Duggan, M. E.; Hartman, G. D.; Hoffman, W. F.; Meissner, R. S.; Perkins, J. J.; Askew, B. C.; Coleman, P. J.; Hutchinson, J. H.; Naylor-Olsen, A. M. U.S. Patent 5,981,546, 1999; For preparation of the radioligand, see: Hamill, T. G.; Duggan, M. E.; Perkins, J. J. *J. Label. Comp. Radiopharm.* **2001**, *44*, 55.
- Marguerie, G. A.; Plow, E. F.; Edgington, T. S. *J. Biol. Chem.* **1979**, *254*, 5357.
- Yasuda, N.; Hsiao, Y.; Jensen, M. S.; Rivera, N. R.; Yang, C.; Wells, K. M.; Yau, J.; Palucki, M.; Tan, L.; Dormer, P. G.; Volante, R. P.; Hughes, D. L.; Reider, P. J. *J. Org. Chem.* **2004**, *69*, 1959.
- Albeck, A.; Persky, R. *J. Org. Chem.* **1994**, *59*, 653.